

CARBON-13 NUCLEAR MAGNETIC RESONANCE SPECTRUM OF COLCHICINE : A REASSIGNMENT

Artur BLADE-FONT^a, Robert MULLER^b, José ELGUERO^c, Robert FAURE^c and Emile-Jean VINCENT^c

^aProductos Frumtost, S.A., Suiza 9-11, Barcelona-6, Spain; ^bDepartement de Chimie Organique, Université de l'Etat à Mons, 7000 Mons, Belgium; ^cLaboratoires de Chimie Moléculaire et de Chimie Organique Physique, Université d'Aix-Marseille III, 13397 Marseille Cédex 4, France

A new NMR signal assignment of the twenty-two carbon atoms of colchicine is proposed that differs considerably from that of Singh, Palmar, Stenberg and Farnum [Spectroscopy Letters, 10, 1001 (1977)]. Some reasons of those discrepancies are analyzed.

The great biological and pharmacological interest of colchicine¹ justifies its study by ¹³C NMR spectroscopy. A correct and complete assignment is furthermore absolutely necessary if the chemical shifts have to be used for a subsequent study of the interaction between colchicine and the tubuline enzyme or for the identification of its metabolites.

Recently, a paper from Singh, Palmar, Stenberg and Farnum² has been published in which the ¹³C NMR spectrum of colchicine in CDCl₃ is given and a complete assignment of the 22 carbon signals proposed. We had in progress a similar study of colchicine and a dozen of its derivatives. The fact that our assignment and that of Singh *et al.*² agree only for 12 of the 22 colchicine carbon atoms, prompts us to publish our assignment and to explain briefly the reasons which in our opinion, have misled the quoted authors.

Like Singh *et al.*² we have recorded the proton-noise-decoupled and the SFORD (single frequency off-resonance decoupled) ¹³C NMR spectra (but in DMSO-d₆ instead of CDCl₃) and compared colchicine C ring to tropolone methyl ether. In addition we have studied many other colchicinic and isocolchicin derivatives and measured the T₁ relaxation times of the different carbon atoms. The results are gathered in Table 1.

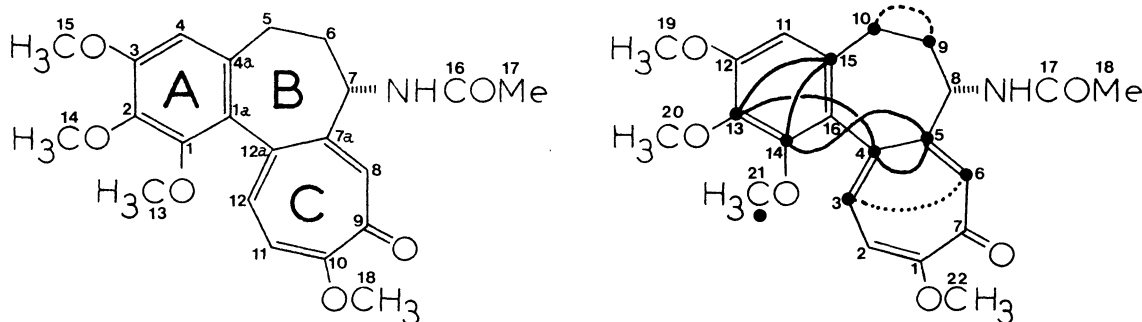


TABLE 1. Carbon-13 chemical shifts of colchicine³ (δ in ppm relative to TMS)

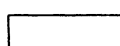
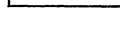
This work:	Carbon No	Assignment	Singh <i>et al.</i> ²	Assignment (No)	Difference
	13	60.9*		56.4 (21)	4.5 ●
	14	60.7*		61.3 (20)	-0.6
	15	56.0**		56.4 (19)	-0.4
	16	168.9		170.3 (17)	-1.4
	17	22.4		22.6 (18)	-0.2
	18	55.9**		56.1 (22)	-0.2

TABLE 1 (Continued)

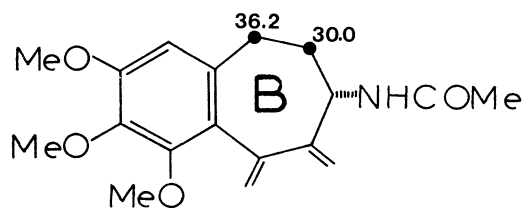
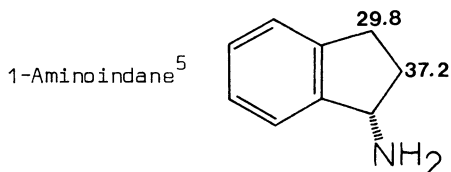
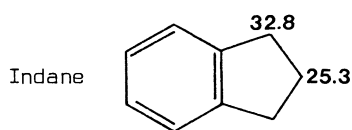
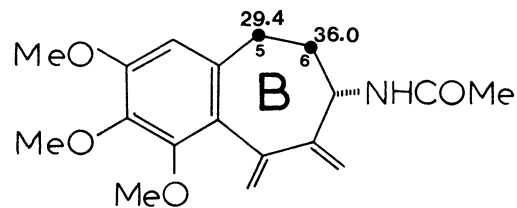
Ring A	1a	125.7		125.7	(16)	0.0
	1	150.7	—	152.9	(14)	-2.2 ●
	2	141.1	—	134.5	(13)	6.6 ●
	3	153.2	—	153.6	(12)	-0.4
	4	108.0	—	107.5	(11)	0.5
	4a	134.4	—	151.1	(15)	-16.7 ●
Ring B	5	29.4	- - -	36.2	(10)	-6.8 ●
	6	36.0	- - -	30.0	(9)	6.0 ●
	7	51.7	- - -	52.9	(8)	-1.2
Ring C	7a	151.2	—	137.2	(5)	14.0 ●
	8	134.7	· · ·	130.5	(6)	4.2 ●
	9	178.4	· · ·	179.6	(7)	-1.2
	10	163.8	· · ·	164.1	(1)	-0.3
	11	112.3	· · ·	113.1	(2)	-0.8
	12	130.7	· · ·	135.8	(3)	-5.1 ●
	12a	135.6	—	141.6	(4)	-6.0 ●

Origin of the mistakes of Singh et al.²Methyl carbon atom, C₁₃.

Due to the lack of resolution (the authors worked at 15.03 MHz instead of 20.0 MHz) they observed only three OCH₃ signals at δ 56.1, 56.4, and 61.3. They supposed the missing signal, corresponding to C₁₃ to be mixed with C₁₅ (56.4), but actually it was under the C₁₄ signal (61.3). That the coincidence of C₁₃ and C₁₄ signals was due to the lower resolution and not to a solvent effect, is proved by the fact that Battersby *et al.*,⁴ in a previous publication, observed in CDCl₃, but not assigned, four signals for the four OCH₃ groups at δ 55.8, 56.1, 61.0, and 61.2.

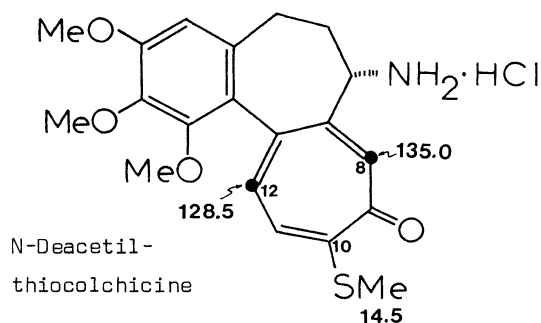
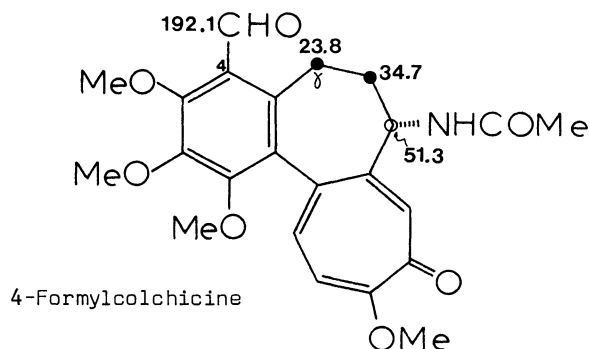
Methylene carbon atoms, C₅ and C₆.

Singh *et al.*² assigned the signals of these two carbon atoms of ring B by comparison with indane. They forgot the β and γ effects produced by an amino group; actually, the comparison with 1-aminoindane⁵ leads to the inverse assignment:

Singh *et al.*² assignment

Our assignment

Moreover, our assignment of C_5 and C_6 signals is not founded on the 1-aminoindane similarities but on the effects produced by a substituent ($CHCl_2$ or CHO) in position 4: steric γ effect on C_5 .

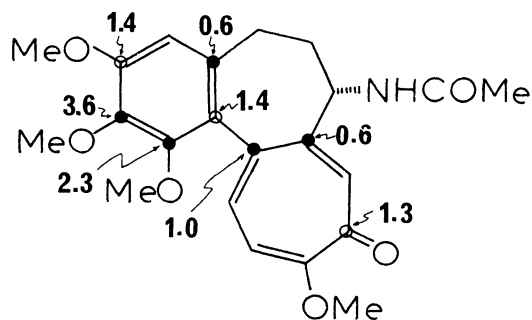
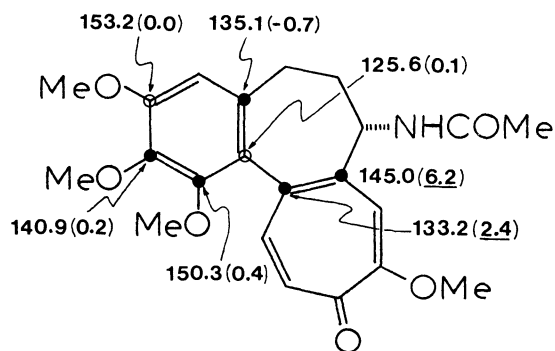


Methine carbon atoms, C_8 and C_{12} .

The assignment of signals at δ 130.5 and 135.8 ($CDCl_3$) by Singh *et al.*² is arbitrary. In fact they are too close to be assigned by comparison with tropolone methyl ether, because of the influence of ring C substituents in positions 7a and 12a. Our assignment, inverted in regard to Singh *et al.*² is supported by the study of N-deacetylthiocolchicine (a derivative with a SCH_3 group instead of a OCH_3 group in position 10).

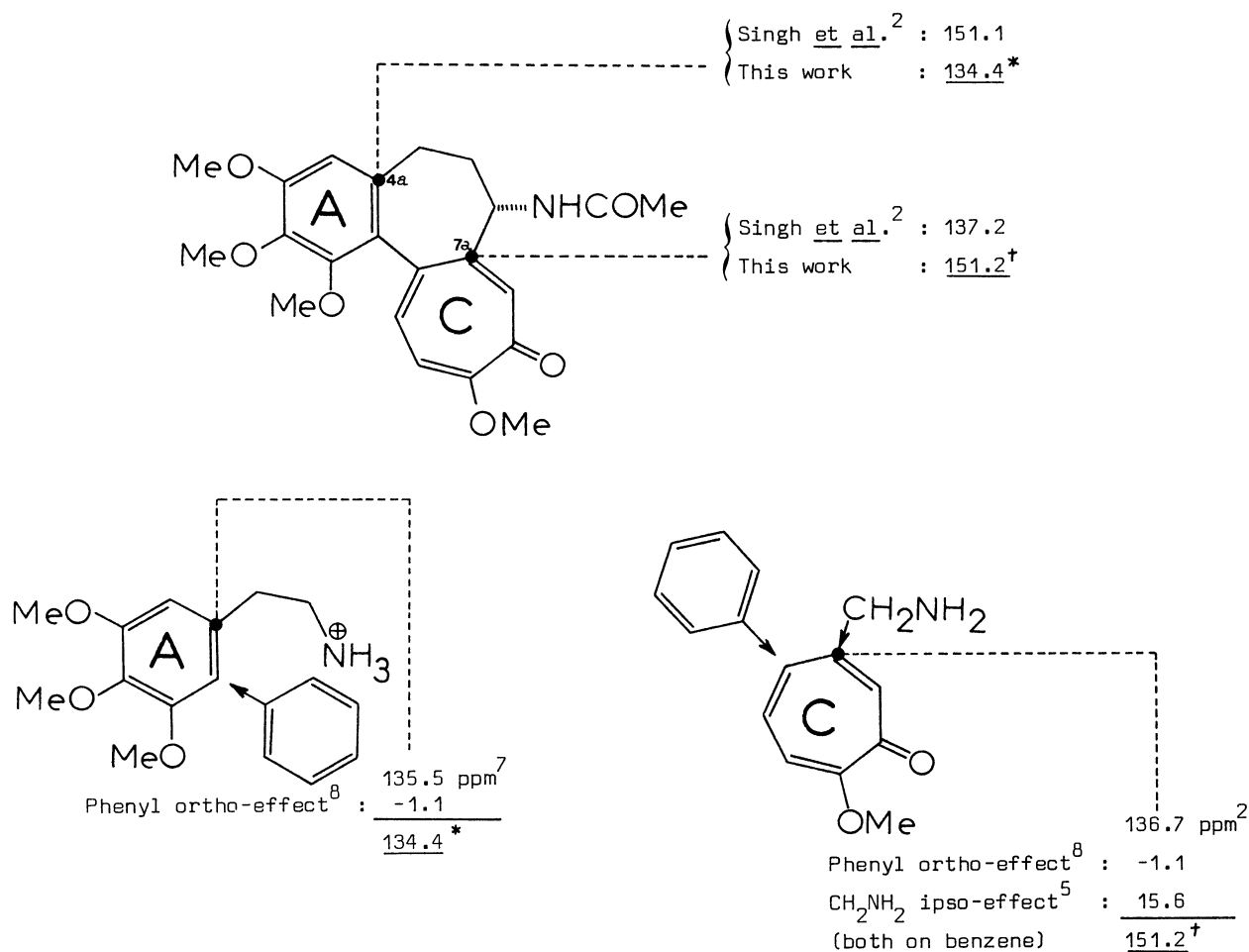
Quaternary carbon atoms, C_1 , C_2 , C_{4a} , C_{7a} , and C_{12a} .

Due to the fact that Singh *et al.*² studied only colchicine itself, they could not differentiate between aromatic quaternary carbon atoms belonging to rings A (benzenic) and C (tropolonic). Because of the study of differently substituted colchicines (substituents in positions 4, 9, and 10 are particularly relevant), we could classify the benzenic and tropolonic carbon atoms into two different groups. After this classification has been done, the assignment is based on T_1 values.^{6,7}



$$(\Delta\delta) = \delta_{\text{isocolchicine}} - \delta_{\text{colchicine}}$$

The most important differences between both assignments are found for quaternary carbon atoms, and amongst them for C_{4a} (-16.7 ppm) and C_{7a} (+14.0 ppm). It is possible to estimate chemical shifts of those two carbons from the literature values of mescaline⁷ and tropolone methyl ether.²



Beyond the accidental coincidence between the experimental and the above calculated values for C_{4a} and C_{7a} carbon atoms, this rough calculations allows no doubt concerning what assignment is the correct one.

REFERENCES

- 1) "The Alkaloids" (R.H.F. Manske, Ed.), Vols. II and VI, Academic Press, New York, N.Y. (1952, 1960); "The Biology of Cytoplasmic Microtubules", Ann. N.Y. Acad. Sci., 253, especially pp. 213-231 (D. Soifer, Ed.).
- 2) S.P. Singh, S.S. Parmar, V.I. Stenberg, and S.A. Farnum, *Spectroscopy Letters*, 10, 1001 (1977).
- 3) Spectrometer Varian CFT-20 working at 20.0 MHz; sample concentration: 0.5 N; tube diameter: 8 mm; spectral width: 4000 Hz; acquisition time: 1.023 sec.; pulse width (flipping angle): 8 sec. (45°); number of data points: 8192; number of transients: 6000.
- 4) A.R. Battersby, P.W. Sheldrake, and J.A. Milner, *Tetrahedron Letters*, 3315 (1974).
- 5) J. Elguero, R. Faure, J. Llinares, and E.J. Vincent, *unpublished results*.
- 6) F.W. Wehrli, *Chem. Commun.*, 379 (1973).
- 7) G.C. Levy, J.D. Cargioli, and F.A.L. Anet, *J. Am. Chem. Soc.*, 95, 1527 (1973).
- 8) J.B. Stothers, "Carbon-13 NMR Spectroscopy", Academic Press, New York, N.Y., p. 90 (1972).

(Received November 27, 1978)